Effect of various sources and levels of iodine, as well as the kind of diet, on the performance of young laying hens, iodine accumulation in eggs, egg characteristics, and morphotic and biochemical indices in blood

M. Słupczyńska,¹ D. Jamroz, J. Orda, and A. Wiliczkiewicz

Department of Animal Nutrition and Feed Science, Wrocław University of Environmental and Life Sciences, 51-630 Wrocław, Poland

ABSTRACT Young hens were fed over a period of 150 d with 2 kinds of diets including corn and soybean meal or corn, soybean, and rapeseed meal. Diets were enriched with potassium iodide (KI) or potassium iodate (KIO_3) as an I source in amounts equal to 1, 3, or 5 mg of supplemented I/kg of feed. The hen performance, egg quality, hematological and morphotic indices in blood, hepatic enzyme activity, lipid indices in blood serum as well as I accumulation in wet egg content were determined. Introduction of 00-variety rapeseed meal into the diet improved the laying rate and feed conversion (P < 0.05); however, better egg weight was noted by feeding the hens with a diet without rapeseed meal. Use of KI as an I source enhanced the egg weight. The increased I level in the diet had an equivocal influence on egg weight, improved the feed conversion per 1 kg of eggs, and decreased the proportion of damaged eggs. The use of corn, soybean, and rapeseed meal in hen diets significantly improved yolk color; similar results were noted after an increase in I levels in the diets after 3 mo of feeding. Hematological indices of hen blood demonstrated significantly higher red blood cells numbers and hemoglobin concentrations with the use of KI. The use of a diet containing rapeseed meal led to an enhancement of hepatic enzyme activity, especially of alkaline phosphatase (P = 0.007). Lipid metabolism indices were not influenced by the kind of diet or the I source or level. The accumulation of I in wet egg content was negatively influenced by the use of a diet containing rapeseed meal (P = 0.000). The application of KI as an I source enhanced (P = 0.003) the accretion of I in eggs after 5 mo of treatment. Enhanced I supply significantly increased accumulation of I in eggs (P = 0.000) after 3 and 5 mo of the experiment from 260 and 310 to 1,011 and 1,256 μ g/kg of wet egg content, respectively.

Key words: hen, iodine, performance, egg, iodine accumulation

2014 Poultry Science 93:2536–2547 http://dx.doi.org/10.3382/ps.2014-03959

INTRODUCTION

The biological necessity of I is mainly related to thyroid gland functions (Van Middlesworth, 1996), especially as a constituent of triiodothyronine and tetraiodothyronine hormones, which play an essential role in metabolism regulation, intermediary cell activity, and cellular oxidation processes (Hetzel and Maberly, 1986; Sourgens et al., 1986; McDowell, 1992; Delange, 1998; Goldhaber, 2003; Lewis, 2004). Moreover, reproduction properties (Trávníček et al., 1997), circulation and muscular systems, maturity processes of cells and tissues, functions of the brain and nervous system, and pituitary gland activity, skin, and secondary skin product formation and regeneration indirectly depend on the supply of I to an organism (Delange, 1994, 1998, 2002; Van Middlesworth, 1996; Liu et al., 2001). Thyroid hormones perform an important role in the regulation of cholesterol levels in humans and rats (Mathe and Chevallier, 1976; Ryś et al., 1997). Excess of this trace element (200–500 mg of I/kg) evokes a dangerous toxicosis in organisms and disorders in the conversion of triiodothyronine to thyroxin (Arrington et al., 1967; Marcilese et al., 1968; Bürgi et al., 2001; Goldhaber, 2003; Baker, 2004).

Plant foods used in human nutrition are generally poor in I, which leads to insufficient I intake and, in consequence, to I deficiency (Haldimann et al., 2005, Röttger et al., 2012). With the aim of increasing I levels in human diets, the application of iodized salt has been popular. However, this is not always a beneficial solution for all human groups (i.e., children, pregnant woman, or the sick; Nair et al., 1998; Szybinski et al., 2001; Brzóska et al., 2005). Iodine may be provided

^{©2014} Poultry Science Association Inc.

Received February 10, 2014.

Accepted June 8, 2014.

¹Corresponding author: maja.slupczynska@up.wroc.pl

in I-enriched milk, fish, and sea products (He et al., 2000; Delange, 2002; Haldimann et al., 2005), but also through fortification of I contents in basic human food (e.g., in eggs; Groppel et al., 1991; Kaufmann et al., 1998, Yalçin et al., 2004).

Enrichment of animal origin products with I could be achieved in a nutritional way by application of various dietary I sources and I levels added into the animal diet. Compounds such as KI, NaI, $Ca(IO_3)_2$, and $Ca(IO_3)_2 \times 6H_2O$ are allowable as I sources. The NRC (1994) recommendations stated the I requirement for young and adult hens at 0.33 to 0.48 mg of I/kg, whereas the German Society for Nutrition Physiology (GfE, 1999) recommendations permit the level of 0.5 mg of I/kg of DM of feed. In EU Commission (2005) regulation 1459/2005 (WHO, UNICEF, ICCIDD, 2001; EFSA, 2005), the permitted I levels for laying hens were reduced from 10 to 5 mg per kg of feed (12%) of humidity). Therefore, the acceptance by ethical commissions of studies in which greater I dietary levels will be used may be impossible.

Absorption and utilization of I from animal diets could be negatively modified by goitrogenic substances that are present in some plant origin feeds, especially those originating from plants of the *Cruciferae* family. The use in animal feed of rapeseed products that has been popular in many countries may decrease the I binding in thyroid hormones and affect thyroid gland functions (Wheeler and Hoffman, 1950; Wemheuer and Paufler, 1993; Tripathi et al., 2001a,b).

The aim of the present study was to examine the effect of various sources and levels of bioactive mineral elements (i.e., I supplements) as well as the presence of rapeseed meal in laying hen diets on the efficacy of egg content enrichment with I. Moreover, performance and egg traits were recorded during performed studies. Because high levels of I were applied for the whole experiment, morphological and biochemical indices in blood and blood serum were assayed as indices of the health status of hens.

MATERIALS AND METHODS

Birds and Management

Sixteen-week-old Hy-Line Brown pullets (1.34 kg of BW) were located in battery pens and before the laying period were fed with a mixture containing about 145 g/ kg of CP and 11.5 MJ/kg of ME.

At the age of about 18 wk, at the beginning of laying, the hens were allocated into cages (3 hens per cage) according to BW and laying at the start time. Afterward, the cages were randomly divided into 12 treatments each consisting of 6 replications (i.e., 18 hens per treatment). The free access of birds to water (nipples) was ensured. The environmental conditions in the summer to autumn period were registered; the room temperature varied between 16 and 25°C and the lighting program was set up for 14 h of light daily. The management was in compliance with European Union regulations. The Local Ethics Commission for Experiments with Animals accepted all experimental procedures.

Diets and Feeding Program

The diet composition was based on our own chemical analyses of feed components and was calculated using linear optimization. The energy concentration was calculated by the use of values given in the European Table of Energy Values for Poultry Feedstuffs (WPSA, 1989). Both prepared basal diets [i.e., cornsoybean mixture (CSM) and corn-soybean-rapeseed meal mixture (CSRM)] were isoproteic and isoenergetic and contained about 165 g/kg of CP and 11.6 MJ of ME/kg. Each diet was enriched with I using either KI or KIO_3 in amounts appropriate to the anticipated I supplement at the level of 1, 3, and 5 mg of I/kg of feed (Table 1). All the feed mixtures were prepared before the experiment. A homogenous mixture of I within the feed was obtained by preparation of premixtures of I supplements with an I free mineral-vitamin premix that was made for the purposes of the experiment. The determined concentration of I in 1 kg of basal diet consisted of 0.52 and 0.49 mg of I/kg of CSM and CSRM, respectively. The feed mixtures were given at a dosage of 110 g per hen daily.

Analytical Methods in Feed Mixtures

The chemical composition of feed components and complete mixtures was determined according to standard methods (AOAC International, 2000). The nitrogen content was determined using a Kjeltec 2300 Foss Tecator apparatus (Höganäs, Sweden) and CP content was calculated by multiplying the nitrogen content by 6.25 (AOAC, 984.13). Crude fiber content was analyzed according to the Henneberg-Stohmann method with the use of a Fibertec Tecator apparatus (Häganäs, Sweden, AOAC, 978.10). After prior wet mineralization of samples with nitric acid (HNO₃) using a MarsX apparatus (CEM Corporation, Matthews, NC), calcium and sodium content in feeds was determined by atomic absorption spectrophotometry using an AA 240 FS apparatus with SIPS 20 (Varian, Mulgrave, Australia; AOAC, 968.08). Phosphorus concentration in feeds was analyzed after prior wet mineralization with nitric acid (HNO_3) and perchloric acid $(HClO_4)$ according to the ammonium vanadomolybdate method using a Specol 11 spectrophotometer (Carl Zeiss, Jena, Germany) at a wavelength of 470 nm (AOAC, 965.17). With the aim of determining amino acid composition, samples of components were hydrolyzed with 6 M hydrochloric acid (HCl) for 24 h at 110°C, then amino acids were separated according to the Moore and Stein (1963) method. For the determination of sulfur amino acids, the feed

	Diet		
Item	CSM	CSRM	
Ingredient, % of diet	·		
Corn meal	15.00	15.00	
Wheat meal	28.44	24.09	
Barley meal	20.00	20.00	
Soybean meal, solvent extracted	20.18	13.23	
Rapeseed 00 meal, solvent extracted		10.00	
Rapeseed oil	4.28	5.71	
Dicalcium phosphate	0.481	0.435	
Chalk	8.931	8.851	
NaCl	0.015	0.013	
DL-Methionine	0.046	0.024	
L-Lvsine	0.128	0.148	
Vitamin and mineral premix ^{2}	2.50	2.50	
Nutritive value of mixtures			
ME, MJ/kg	11.65	11.65	
CP, g/kg	165.00	162.00	
Crude fiber, g/kg	36.26	28.97	
Ca. g/kg	38.80	38.90	
P available, g/kg	4.30	4.32	
Na, g/kg	1.70	1.71	
Methionine, g/kg	4.30	4.31	
Lysine, g/kg	9.00	8.97	
I, mg/kg	0.52	0.49	
Nutritive recommendation for Hy-Line laying hens			
ME, MJ/kg	11	.65	
CP. g/kg	165	5.00	
Crude fiber, g/kg	to 40	0.00	
Ca. g/kg	38	3.80	
P available, g/kg	4	1.30	
Na. g/kg	1	.70	
Methionine, g/kg	2	1.30	
Lysine, g/kg	(0.00	

 Table 1. Basal composition of the experimental diets¹

 $^1\mathrm{CSM}=\mathrm{corn}\text{-soybean}$ mixture; $\mathrm{CSRM}=\mathrm{corn}\text{-soybean}\text{-rapeseed}$ meal mixture.

²Supplied per kilogram of diet: vitamin A, 12,000 IU; vitamin D₃, 3,300 IU; vitamin E, 30.0 mg; vitamin K₃, 3.0 mg; vitamin B₁, 2.20 mg; vitamin B₂, 6.60 mg; vitamin B₆, 3 mg; vitamin B₁₂, 27 μ g; pantothenic acid, 10.2 mg; nicotinic acid, 30 mg; folic acid, 0.9 mg; choline, 0.315 mg; Mg, 113 mg; Zn, 72 mg; Co, 0.48 mg; Cu, 10.0 mg; Fe, 60 mg; Se, 0.30 mg; I, 0.0 mg.

samples were oxidized (0°C, 16 h) with formic acid and hydrogen peroxide (H₂O₂; 9:1, vol:vol) before HCl hydrolysis and then were separated using an AAA 400 Ingos Analyzer (Prague, Czech Republic). The I content in basal mixtures was analyzed by a spectrophotometric method based on the Sandell-Kolthoff reaction (Gasior and Szczypuła, 2010).

Performance, Egg Traits—Data Collection

Data collection was realized for very young hens with the aim of the designation of their reaction to I supply over the period of the beginning of egg production.

The laying performance of young hens was recorded over a period of 150 d. The number of laying eggs, from 10 d after the start of laying, was registered for each cage daily and all the eggs were weighed individually. Moreover, the number of damaged eggs were recorded. Feed intake and the mortality of hens were controlled weekly for each cage-replication. On the basis of the collected data mean laying rates (%), egg weight (g), egg mass (g/head per day), feed efficiency per egg (g/g), and per kilogram of eggs (kg/kg), as well as the number of damaged eggs (%) were calculated for the whole period of the experiment.

After 3 and 5 mo of feeding with the experimental diets, 30 eggs from each treatment were randomly sampled. Collected eggs were weighed as well as the yolk, albumen, and eggshell. Eggshell thickness (mm) was measured with a micrometer screw (0.001-mm micrometer IP 54, Wilson Wolpert, Maastricht, the Netherlands). Eggshell breaking strength (kg) was measured using a ZWICK/Roell (Ulm, Germany) apparatus. Each measurement was repeated twice. Egg yolk color intensity was evaluated visually using a La Roche yolk color fan scale.

Determination of lodine Content in Eggs

At the end of 3 and 5 mo of the experiment, 12 eggs were randomly sampled from each treatment. Their content was freeze-dried and mineralized at 580°C with NaOH + ZnSO₄ × 7H₂O supplement. Iodine concentrations in egg content were determined according to the method modified by Gąsior and Szczypuła (2010), on the basis of the speed of reaction catalyzed by I: 2

Table 2. Laying hen performance¹

				Feed efficiency			
Item	Laying rate, %	Egg weight, g	Egg mass, g/hen per day	Per egg, g	Per kg of eggs, kg	$\begin{array}{c} \text{Damaged} \\ \text{eggs}, \ \% \end{array}$	
Treatment							
CSM	0.0	50.0		100	0.10	r ooA a	
1 mg of I as KI	83.0	56.0	45.5	122	2.18	5.20 ^{11,a}	
3 mg of I as KI	77.5 ^{11,0}	56.6	43.8 ^{11,a}	132 ^{11,a}	$2.32^{11,\alpha}$	2.23 ^b	
5 mg of I as KI	87.35	57.3 ^{rr}	50.12	1165	2.025	2.375	
1 mg of 1 as KIO_3	83.8	55.75	46.6	121 114 B	2.17 2.04 B	3.58	
$3 \text{ mg of I as KIO}_3$	88.4	56.0	49.5	114 5	2.04	2.53	
5 mg of 1 as KIO ₃ CSRM	85.5	56.3	48.1	118	2.10	2.045	
1 mg of I as KI	86.0	56.6	48.7	118^{b}	2.08	4.02	
3 mg of I as KI	85.1	56.0	47.6	119	2.13^{b}	1.69^{B}	
5 mg of I as KI	88.8^{B}	55.6^{B}	49.4^{b}	114^{B}	2.04^{B}	1.50^{B}	
$1 \text{ mg of I as KIO}_3$	87.2^{b}	56.0	48.8^{b}	116^{b}	2.07^{b}	4.39	
$3 \text{ mg of I as KIO}_3$	86.1	55.0^{B}	47.3	117^{b}	2.14	2.61	
$5 \text{ mg of I as KIO}_3$	86.2	56.7	48.8^{b}	117^{b}	2.07	3.16	
SEM	0.605	0.103	0.337	0.001	0.016	0.201	
<i>P</i> -value	0.009	0.000	0.006	0.004	0.004	0.000	
Kind of diet							
CSM	84.2 ^a	56.3^{a}	47.4	120^{a}	2.13	2.99	
CSRM	86.6^{b}	56.0^{b}	48.4	116^{b}	2.09	2.89	
I source							
KI	84.6	56.4^{a}	47.7	120	2.13	2.83	
KIO ₃	86.2	55.9^{b}	48.2	117	2.10	3.05	
Level of I supplement, mg							
1	85.0	56.1	47.7	119	2.13^{ab}	4.29^{A}_{-}	
3	84.3	55.9^{a}	47.0^{a}	121	2.16^{a}	2.26^{B}_{-}	
5	87.0	56.5^{b}	49.1^{b}	116	2.06^{b}	2.27^{B}	
Source of variation, <i>P</i> -value							
Kind of diet	0.035	0.038	0.093	0.026	0.067	0.778	
I source	0.157	0.011	0.389	0.108	0.288	0.520	
I level	0.120	0.014	0.020	0.106	0.021	0.000	
Kind of diet \times I source	0.112	0.131	0.200	0.082	0.145	0.026	
Kind of diet \times I level	0.709	0.006	0.346	0.636	0.363	0.896	
I source \times I level	0.011	0.125	0.031	0.010	0.027	0.212	
Kind of diet \times I source \times I level	0.111	0.006	0.036	0.090	0.036	0.632	

^{a,b}Means within a column marked with different superscripts differ significantly at P < 0.05.

^{A,B}Means within a column marked with different superscripts differ significantly at P < 0.01.

 1 CSM = corn-soybean mixture; CSRM = corn-soybean-rapeseed meal mixture.

 $\mathrm{Ce}^{4+} + \mathrm{As}^{3+} \rightarrow 2 \ \mathrm{Ce}^{3+} + \mathrm{As}^{5+}$ (Knapp et al., 1998; Zhang et al., 2005). The principle of the assessment is the reduction of Ce^{4+} to Ce^{3+} in the presence of As^{3+} , where iodide ions (I⁻) act as a catalyst for this reaction.

Blood Collection and Analysis

At the end of the experiment, 6 hens were randomly selected from each treatment and designated for blood sampling. Blood samples were taken from the vena brachialis by a specialized veterinarian and divided into 2 test tubes. In whole heparinized blood samples, the health status hematological indices were determined: red blood cells (**RBC**), hemoglobin (**HGB**), hematocrit, platelets, and leukogram—white blood cells, as well as the proportions of lymphocytes, heterophiles, monocytes, basophils, and eosinophils. Moreover, the activity of hepatic enzymes was assayed in blood serum: aspartate aminotransferase (**AST**), alanine aminotransferase (**ALT**), and alkaline phosphatase (**ALP**). The same was performed for indices of lipid metabolism: triglycerides, total cholesterol, high-density lipoproteins, and low-density lipoproteins. The hematological indices were assayed by the optic and impedance method using a Cell-Dyn 3700 (Abbott Laboratories, Abbott Park, IL). In blood smears colored according to the May–Grunwald–Giemza method, leukograms were microscopically determined by the use of a Burker chamber and Natt and Harric liquid. Biochemical indices in blood serum were determined spectrophotometically by the use of an XL300, Erba apparatus, and Biosystem chemicals. All blood analyses were conducted at Vet-Lab, a certified veterinary laboratory in Wroclaw.

Statistical Evaluation of Results

All numerical data were evaluated using 1-, 2-, or 3 factorial ANOVA using Statistica version 10 computer software (Statistica, StatSoft Inc., 2012). The differences among treatments and for experimental factors were tested according to the following statistical models:

$$y_{ij} = \mu + \alpha_i + e_{ij}$$

(for differences between treatments),

$$y_{jik} = \mu + \alpha_i + \beta_j + (\alpha\beta)_{ij} + e_{ijk}$$

(for kinds of diet or inclusion levels of I supplement

or their levels), or

$$y_{ijkl} = \mu + \alpha_i + \beta_j + \gamma_k + (\alpha\beta)_{ij} + (\alpha\gamma)_{jk} + (\alpha\beta\gamma)_{ijk} + e_{ijkl},$$

where y_{ij} , y_{ijk} , and y_{jikl} are the variance associated with parameter α ; μ is the overall mean; α_i is the treatment effect; β_j is the kind of diet, I-supplement, or their level; γ_k is an effect for the iodine level; $(\alpha\beta)_{ij}$ is the interaction effect; and e_{ij} or e_{ijk} (e_{ijkl}) is an error term.

The individual measurements or averages for replication laying rates or feed consumption were treated as experimental units, and differences between treatment averages (means) were analyzed for significance (P < 0.05 or P<0.01) using Tukey's multiple-range test. All numerical data in tables are presented as average values and are accompanied by SEM values.

RESULTS AND DISCUSSION

Hen Performance

The productivity of young hens was acceptable. The laying rate over the 150 d of the experimental period was a mean of about 85%, with an average egg weight of about 56 g. Hen performance was affected by experimental factors (Table 2). Hens receiving 10% rapeseed meal in their diet demonstrated higher laying rates (P = 0.035) and better feed efficiency per egg (P = 0.026), but lower egg weights (P = 0.038) compared with those in hens receiving rations without rapeseed meal. Thus, it could be stated that incorporation of 10% rapeseed meal into the feed mixture for hens had a positive effect on hen performance. Gheisari et al. (2011), in experiments with laying hens, stated that including different levels (0, 5, 10, 15, and 20%) of rapeseed meal in the

Table 3. Effect of experimental factors on egg characteristics after 3 mo of treatment¹

	Proportion of egg weight, $\%$			Yolk color,	Shell	Shell
Item	Yolk	Albumen	Shell	points	kg	mm
Treatment						
CSM						
1 mg of I as KI	29.9	56.4	13.3	12.7	3.45	0.354
3 mg of I as KI	27.8	57.9	14.1	11.5 ^A	3.95	0.347
5 mg of I as KI	30.3	56.5	14.2	$13.0^{BCD}_{}$	3.70	0.344
1 mg of I as KIO ₃	30.6	55.5	14.3	13.2^{BCD}	3.97	0.372
$3 \text{ mg of I as KIO}_3$	30.9	54.2	14.9 ^A	12.7	3.42	0.376
$5 \text{ mg of I as KIO}_3$	30.1	55.2	14.7	12.2^{B}	4.07	0.380
CSRM				120		
1 mg of I as KI	29.5	57.1	14.0	$12.5^{\text{ABC}}_{\text{D.GD}}$	3.47	0.355
3 mg of I as KI	30.4	55.5	14.2	$13.2^{\text{BCD}}_{\text{D}}$	3.52	0.358
5 mg of I as KI	29.4	56.1	14.5	14.0 ^D	3.35	0.372
1 mg of I as KIO ₃	29.1	56.1	14.1	$12.5^{\text{ABC}}_{}$	3.77	0.351
$3 \text{ mg of I as KIO}_3$	28.0	58.0	14.3_	13.0^{BCD}	3.95	0.353
$5 \text{ mg of I as KIO}_3$	28.8	58.1	13.0^{B}	13.7^{CD}	3.85	0.349
SEM	0.236	0.298	0.110	0.113	0.068	0.003
P-value	0.096	0.102	0.011	0.000	0.304	0.266
Kind of diet						
CSM	29.9	55.8	14.2	12.6^{A}_{-}	3.76	0.362
CSRM	29.1	56.8	14.0	13.2^{B}	3.65	0.356
I source						
KI	29.5	56.6	14.1	12.8	3.57	0.355
KIO ₃	29.5	56.1	14.2	12.9	3.84	0.363
Level of I supplement, mg						
1	29.7	56.2	13.9	12.7^{a}	3.67	0.357
3	29.3	56.4	14.4	$12.6^{A,a}$	3.71	0.358
5	29.6	56.5	14.1	$13.2^{\mathrm{B,b}}$	3.74	0.362
Source of variation, <i>P</i> -value						
Kind of diet	0.095	0.094	0.251	0.000	0.418	0.398
I source	0.945	0.387	0.404	0.577	0.051	0.194
I level	0.647	0.882	0.177	0.003	0.897	0.863
Kind of diet \times I source	0.012	0.005	0.002	0.100	0.291	0.004
Kind of diet \times I level	0.630	0.916	0.130	0.000	0.583	0.893
I source \times I level	0.758	0.498	0.040	0.024	0.252	0.878
Kind of diet \times I source \times I level	0.036	0.095	0.282	0.032	0.192	0.464

^{a,b}Means within a column marked with different superscripts differ significantly at P < 0.05.

^{A–D}Means within a column marked with different superscripts differ significantly at P < 0.01.

 1 CSM = corn-soybean mixture; CSRM = corn-soybean-rapeseed meal mixture.

IODINE ADDITIVES IN HENS

Table 4. Effect of e	xperimental	factors on e	egg characteristics	after 5 m	o of treatment ^{1}
----------------------	-------------	--------------	---------------------	-----------	--

	Prope	Proportion of egg weight, $\%$			Shell	Shell
Item	Yolk	Albumen	Shell	points	strength, kg	mm
Treatment						
CSM						
1 mg of I as KI	31.6	54.6	13.8^{A}	12.7	4.25	0.369
3 mg of I as KI	31.3	53.9	14.4	13.5	4.02	0.371
5 mg of I as KI	31.9	53.1	14.9	12.2	3.82	0.372
$1 \text{ mg of I as KIO}_3$	32.9	52.0	15.2	13.0	4.17	0.379
$3 \text{ mg of I as KIO}_3$	33.1	51.1^{a}	14.4^{A}	12.7	4.10	0.369
$5 \text{ mg of I as KIO}_3$	32.6	52.3	15.1	12.5	3.65	0.347
CSRM						
1 mg of I as KI	32.3	52.3	15.0	13.0	4.05	0.379
3 mg of I as KI	33.9	50.7^{a}	15.3	12.7	3.70	0.347
5 mg of I as KI	29.9	55.2	15.2	13.0	3.70	0.360
$1 \text{ mg of I as KIO}_3$	32.7	52.0	15.3	13.2	3.27	0.350
$3 \text{ mg of I as KIO}_3$	32.1	52.1^{b}	16.0^{B}	13.5	3.65	0.367
$5 \text{ mg of I as KIO}_3$	31.4	54.5	14.3^{A}	13.5	3.47	0.354
SEM	0.238	0.289	0.107	0.091	0.069	0.002
P-value	0.069	0.006	0.000	0.056	0.072	0.051
Kind of diet						
CSM	32.2	52.5	14.6^{A}	12.7^{a}	4.00^{A}	0.367
CSRM	32.0	52.8	15.2^{B}	13.1^{b}	3.64^{B}	0.360
I source						
KI	31.8	53.3	14.8	12.8	3.92	0.366
KIO ₃	32.5	52.3	15.0	13.0	3.72	0.361
Level of I supplement, mg						
1	32.4	52.7	14.8	13.0	3.93	0.369
3	32.6	51.9^{A}	15.0	13.1	3.86	0.363
5	31.4	53.8^{B}	14.8	12.8	3.66	0.358
Source of variation, <i>P</i> -value						
Kind of diet	0.652	0.945	0.001	0.029	0.006	0.104
I source	0.126	0.052	0.117	0.214	0.112	0.240
I level	0.075	0.012	0.502	0.309	0.190	0.162
Kind of diet \times I source	0.176	0.027	0.125	0.085	0.252	0.877
Kind of diet \times I level	0.083	0.009	0.000	0.097	0.432	0.687
I source \times I level	0.562	0.781	0.012	0.643	0.372	0.097
Kind of diet \times I source \times I level	0.120	0.221	0.031	0.152	0.518	0.013

a,b Means within a column marked with different superscripts differ significantly at P < 0.05.

 ${}^{\rm A,B}\!{\rm Means}$ within a column marked with different superscripts differ significantly at P < 0.01.

 1 CSM = corn-soybean mixture; CSRM = corn-soybean-rapeseed meal mixture.

diet induced no effects on their performance, including laying rate, egg weight, and feed conversion ratio. The weight of eggs from hens receiving KI as an I source was higher (P = 0.011) than that observed when using KIO_3 . The I source did not influence the remaining performance results. Kaufmann et al. (1998) also found similar effect of I additives on hens performance results. Opaliński et al. (2012), in experiments with Hy-Line Brown hens, stated that an I source given at the same level, 1 mg/kg, as $Ca(IO_3)_2$ or I-enriched yeast had no effect on estimated hen performance, including egg weight. Higher levels of added I significantly modified the egg weight as well as egg mass (P = 0.014 and P =0.020, respectively), improved the feed efficiency per kilogram of eggs (P = 0.021), and reduced the number of damaged eggs (P < 0.01). Similarly, Saki et al. (2012) reported that increasing I supplementation led to higher egg weights. Contrasting results were obtained by Röttger et al. (2012) who stated that different I concentrations in feed did not affect hen performance. Better results in terms of BW gain, feed intake, and conversion were observed by He et al. (2002) during supplementation of pig diets with KI. These and our results are still in contrast to the findings of Yalçin et al. (2004) and Röttger et al. (2012); however, by enhancing I dosages to 12 and 24 mg/kg, the egg weight was less than that with a lower I supply. Neither these authors nor Richter (1995) stated any significant effect on hen performance.

It is difficult to present unequivocal conclusions concerning the application of either various levels of I supplements or types of diet due to interactions between experimental factors. In hens receiving a CSM diet, the highest laying rate was stated in groups receiving 3 mg of I/kg of a diet given as KIO₃. This was significantly higher (P = 0.009) than in birds on a diet with the same level of I but given as KI where the laying rates in this group of birds were the lowest. Generally, taking into consideration the tendency in hen laying rate regardless of dietary type and at I level of 1 or 3 mg/ kg of diet, better results were obtained if KIO₃ rather than KI served as the I source. The opposite result was observed if the I level was at 5 mg/kg of diet; in that case a slightly higher hen laying rate was stated if the I source was KI. The interaction kind of diet \times I source \times I level was significant (P = 0.006) for egg weight and for feed efficiency (P = 0.036). A similar tendency was

Table 5	Hemato	logical	indices	of	hen	$blood^1$
---------	--------	---------	---------	----	-----	-----------

Itam	RBC,	HGB,	HCT,	PT,
	10 /L		L/L	g/L
Treatment				
CSM				
1 mg of I as KI	2.627	7.40	0.313	53.88
3 mg of I as KI	2.738	7.75	0.325	45.73
5 mg of I as KI	2.640	7.44	0.311	54.65
1 mg of I as KIO ₃	2.450	7.53	0.314	48.65
$3 \text{ mg of I as KIO}_3$	2.345	7.30	0.303	52.95
$5 \text{ mg of I as KIO}_3$	2.242	7.36	0.305	50.80
CSRM				
1 mg of I as KI	2.565	7.39	0.308	50.82
3 mg of I as KI	2.495	7.36	0.311	47.63
5 mg of I as KI	2.457	7.07	0.300	43.95
1 mg of I as KIO ₃	2.428	7.29	0.307	53.32
$3 \text{ mg of I as KIO}_3$	2.403	7.16	0.299	45.92
$5 \text{ mg of I as KIO}_3$	2.395	6.96	0.286	45.00
SEM	0.033	0.090	0.004	1.016
<i>P</i> -value	0.169	0.972	0.943	0.311
Kind of diet				
CSM	2.517	7.46	0.31	51.11
CSRM	2.461	7.20	0.30	47.77
I source				
KI	2.587^{A}	7.40^{A}	0.311	49.44
KIO ₃	2.377^{B}	7.27^{B}	0.302	49.41
Level of I supplement, mg				
1	2.518	7.40	0.310	51.67
3	2.495	7.39	0.309	48.06
5	2.433	7.21	0.300	48.60
Source of variation, <i>P</i> -value				
Kind of diet	0.441	0.554	0.400	0.100
I source	0.002	0.003	0.281	0.998
I level	0.542	0.627	0.563	0.291
Kind of diet \times I source	0.083	0.077	0.987	0.759
Kind of diet \times I level	0.882	0.905	0.903	0.184
I source \times I level	0.840	0.814	0.819	0.625
Kind of diet \times I source \times I- level	0.589	0.638	0.709	0.200

 ${}^{\rm A,B}\!{\rm Means}$ within a column marked with different superscripts differ significantly at P < 0.01.

 1 RBC = red blood cells; HGB = hemoglobin; HCT = hematocrit; PT = platelets; CSM = corn-soybean mixture; CSRM = corn-soybean-rapeseed meal mixture.

shown for feed efficiency calculated in grams per egg. In the experiment conducted by Lichovnikova and Zeman (2004), neither an increased rapeseed meal content in the diet nor I level had any effect on estimated hen performance.

Egg Quality Characteristics

Egg quality characteristics were stated after 3 and 5 mo of the experiment (Tables 3 and 4). After 3 mo of treatment, there was no significant influence as a result of the kind of diet, I source, or I level in the volk, albumen, and shell proportions in the egg weight or in shell strength and thickness (Table 3). Statistical analysis showed significant differences in yolk color determined according to the Roche scale that were higher for volks in eggs of hens receiving CSRM. In eggs of hens receiving for 3 mo in ration I supplements at a level of 1 or 3 mg/kg, yolk color estimated in Roche points was lower than in hens receiving 5 mg/kg of I (P < 0.01). After 5 mo of the experiment, a similar tendency was demonstrated for the effects of the kind of diet, I sources, and I levels on egg characteristics (Table 4). Additionally, the effect of dietary type on the ratio of the eggshell weight to whole egg weight and eggshell strength was shown. Obtained data are difficult to unambiguously interpret due to different kind of interactions between experimental factors. No impact on the eggshell index, eggshell breaking strength, or eggshell thickness as well as the egg yolk index was reported by Yalçin et al. (2004) in an experiment in which the I supplementation was increased from 3 to 24 mg/kg. Opaliński et al. (2012) found higher albumen weight in eggs by application of increasing amounts of I supplements (from 1 to 2 mg of I per kg of feed). Increasing egg weight and eggshell ratio were obtained in treatments with 10 or 15 mg/kg dietary I by Saki et al. (2012); however, the lowest value of albumen height was indicated after supply of 5 mg of I/kg.

=

Hematological and Morphotic Indices in Blood

Diet type and level of I supplement had no effect on the analyzed indices (Table 5). However, unexpected significant (P = 0.002) influences of I sources on RBC and HGB concentrations were found. Values of these indices were higher in blood of hens fed with a diet enriched with I by the use of KI rather than a KIO₃ supplement. All interactions among experimental factors were nonsignificant. The nonsignificant influence of diet type, I source, and I level on white blood cell number and their appearance was shown (Table 6); only the percentage share of monocytes in white blood cells was higher (P = 0.014) in the blood of hens fed with a diet containing soybean meal as a protein source. All analyzed blood indices were in the range of recommendations acknowledged as physiological. All interactions were nonsignificant.

In analyzed publications, only a little information was found concerning the influence of I supply on hematological indices in laying hens. Lewis (2004) reported that applying I supplementation could increase HGB levels in blood. In our studies, a similar effect was found with an I source. Trávníček et al. (2000) reported that a moderate excess (15 mg/kg) of dietary I could lead to a decrease in phagocytic heterophil activity and the phagocytic index, as well as an increase in heterophils in relation to lymphocytes. Nevertheless, the reason for these results cannot be explained and discussed in the context of the available literature.

Hepatic Enzyme Activity in Blood Serum

The mean activity of AST amounted to about 205 to 315 U/L and that of ALT to about 2.7 to 5.9 U/L. The activity of ALP was in total high and varied in a wide range from about 260 to 735 U/L of blood serum (Table 7). The implementation of rapeseed meal into the diet led to a significant (P = 0.07) increase in ALP activity. Other experimental factors did not affect the activities of the remaining hepatic enzymes.

As revealed by Yue and Kang (1995), a high intake of I (65 and 130 mg/kg) from diets evokes elevated plasma aspartate aminotransferase and alanine aminotransferase activity in hens as well as increased activity of alkaline phosphatase. However, unclear or no changes were stated in terms of the activity of other enzymes (i.e., superoxide dismutase, glutamic-pyruvic transaminase). High levels of I in a diet could affect alkaline phosphatase activity (Yue and Kang, 1995). The AST activity determined in our investigation generally shows elevated values as (average about 205–315 U/L) compared with values acknowledged as physiologically correct (142–219 U/L, Krasnodębska-Depta, 2005), but experi-

Table 6. Leukogram of laying hen blood¹

		Percentage in WBC					
Item	WBC, $10^9/L$	Lymphocytes	Heterophils	Monocytes	Basophils	Eosinophils	
Treatment							
CSM							
1 mg of I as KI	14.44	59.33	30.00	7.50	2.33	0.83	
3 mg of I as KI	14.22	65.33	22.50	8.67	2.17	1.33	
5 mg of I as KI	18.33	70.83	20.50	4.50	2.83	1.33	
$1 \text{ mg of I as KIO}_3$	18.91	68.33	23.50	5.67	2.00	0.50	
$3 \text{ mg of I as KIO}_3$	20.10	64.17	25.50	5.17	3.17	2.00	
5 mg of I as KIO_3	16.38	66.17	25.33	4.83	2.33	1.33	
CSRM							
1 mg of I as KI	20.66	69.00	25.83	2.17	1.17	1.83	
3 mg of I as KI	18.67	71.17	21.17	5.17	1.33	1.17	
5 mg of I as KI	14.00	72.17	21.67	3.33	2.67	0.17	
$1 \text{ mg of I as KIO}_3$	21.43	67.50	26.00	3.50	1.67	1.33	
$3 \text{ mg of I as KIO}_3$	21.20	66.67	26.33	5.33	0.67	1.00	
5 mg of I as KIO_3	16.00	71.50	21.33	3.83	3.00	0.33	
SEM	0.869	1.611	1.279	0.440	0.216	0.162	
P-value	0.643	0.958	0.974	0.229	0.423	0.324	
Kind of diet							
CSM	17.06	65.69	24.56	6.06^{a}	2.47	1.22	
CSRM	18.66	69.67	23.72	3.83^{b}	1.75	0.97	
I source							
KI	16.69	67.97	23.61	5.17	2.08	1.11	
KIO ₃	18.94	67.39	24.67	4.72	2.14	1.08	
Level of I supplement, mg							
1	18.78	66.04	26.33	4.63	1.79	1.13	
3	18.43	66.83	23.88	6.08	1.83	1.38	
5	16.27	70.17	22.21	4.13	2.71	0.79	
Source of variation, <i>P</i> -value							
Kind of diet	0.371	0.211	0.608	0.014	0.139	0.582	
I source	0.203	0.985	0.780	0.422	0.970	0.934	
I level	0.407	0.622	0.460	0.286	0.126	0.243	
Kind of diet \times I source	0.772	0.779	0.913	0.275	0.872	0.677	
Kind of diet \times I level	0.277	0.955	0.995	0.404	0.164	0.022	
I source \times I level	0.626	0.727	0.646	0.551	0.933	0.468	
Kind of diet \times I source \times I level	0.612	0.696	0.683	0.668	0.456	0.808	

^{a,b}Means within a column marked with different superscripts differ significantly at P < 0.05.

 1 WBC = white blood cells; CSM = corn-soybean mixture; CSRM = corn-soybean-rapeseed meal mixture.

2543

Item	AST	ALT	ALP
Treatment			
CSM			
1 mg of I as KI	262.3	3.95	349.23^{A}
3 mg of I as KI	215.4	2.75	535.42
5 mg of I as KI	234.4	4.68	$259.72^{A,a}$
1 mg of I as KIO ₃	222.4	4.15	380.72^{A}
$3 \text{ mg of I as KIO}_3$	199.0	5.42	368.02^{A}
$5 \text{ mg of I as KIO}_3$	205.2	3.98	416.20^{a}
CSRM			
1 mg of I as KI	204.9	3.72	385.60^{A}
3 mg of I as KI	204.4	4.07	522.93^{bc}
5 mg of I as KI	259.7	3.47	449.12^{a}
1 mg of I as KIO ₃	218.8	3.43	$598.07^{ m bc}$
$3 \text{ mg of I as KIO}_3$	315.3	5.93	396.67^{a}
$5 \text{ mg of I as KIO}_3$	259.8	4.87	$735.05^{B,b}$
SEM	9.44	0.243	25.90
P-value	0.340	0.339	0.010
Kind of diet			
CSM	223.1	4.16	384.88^{A}
CSRM	243.9	4.25	514.57^{B}
I source			
KI	230.2	3.77	417.00
KIO ₃	236.8	4.63	482.45
Level of I supplement			
1 mg	227.15	3.81	428.40
3 mg	233.54	4.54	455.76
5 mg	239.83	4.25	465.02
Source of variation, <i>P</i> -value			
Kind of diet	0.271	0.849	0.007
I source	0.726	0.079	0.165
I level	0.858	0.463	0.801
Kind of diet \times I source	0.065	0.778	0.213
Kind of diet \times I level	0.155	0.466	0.106
I source \times I level	0.312	0.119	0.006
Kind of diet \times I source \times I level	0.540	0.406	0.825

Table 7. Hepatic enzyme activity $(U/L)^1$

 $^{\rm a-c}{\rm Means}$ within a column marked with different superscripts differ significantly at P<0.05.

 $^{\rm A,B}$ Means within a column marked with different superscripts differ significantly at P < 0.01.

 1 CSM = corn-soybean mixture; CSRM = corn-soybean-rapeseed meal mixture; AST = aspartate aminotransferase; ALT = alanine aminotransferase; ALP = alkaline phosphatase.

mental factors have not significantly modified these indices. The ALT activity was in compliance with physiological standards (Krasnodębska-Depta, 2005) and was not modified by experimental factors. The activity of alkaline phosphatase showed great diversification from about 260 to 735 U/L and exceeded the recommended range of 184 to 458 U/L. The I supply has not clearly influenced the liver function expressed in terms of the activity of selected enzymes. A similar lack of effect on AST, ALT, and ALP activity of I application in pigs was presented by He et al. (2000, 2002).

Lipid Indices in Blood Serum

Lipid metabolites that were determined in the blood serum of young hens were corrected for healthy hens. The triacylglyceride concentration varied from 8.2 to 10.5 mmol/L (Table 8) and the cholesterol level was within the range 2.8 to 4.8 mmol/L, high-density lipoprotein and low-density lipoprotein fractions were approximately 1.5 to 2.3 and 0.9 to 1.7 mmol/L, respectively. Along with the kind of diet, I supplement and I level did not affect the lipid indices in blood serum. All differences among means for the treatments and experimental factors were nonsignificant. The assayed indices are accurate for laying hens (Krasnodębska-Depta, 2005).

These results are in agreement with the findings of He et al. (2002) that the supply of I, even at higher levels (5–8 mg/kg feed), did not affect the triacylglyceride and cholesterol concentrations in blood serum. Similar to the obtained data, no significant differences among treatments in blood serum cholesterol were obtained by the use of 12 or 24 mg of I supply in the Yalçin et al. (2004) experiment. However, as a result of the application of very high I supplements (up to 500–3,000 mg/ kg of feed), dynamically enhanced plasma cholesterol (to about 12 mmol/L) was obtained in the studies of Perry et al. (1989, 1990). Increasing cholesterol levels in serum as a result of high I content in hen diets were also highlighted by Krupová et al. (1998).

Iodine Accumulation in Wet Egg Content

Iodine concentration in wet egg content was determined twice, after 3 and 5 mo of the experiment. Iodine accumulation in egg content was negatively affected by rapeseed meal addition into feed mixture in both sea-

Table 8. Lipid metabolites in laying hens blood serum $(mmol/L)^{1,2}$

	``````````````````````````````````````			
Item	TG	CHOL	HDL	LDL
Treatment				
CSM				
1 mg of I as KI	9.85	4.35	1.68	1.26
3 mg of I as KI	8.26	2.90	1.48	0.88
5 mg of I as KI	9.59	4.02	1.92	1.38
1 mg of I as KIO ₃	9.91	3.75	1.87	1.29
$3 \text{ mg of I as KIO}_3$	10.56	4.85	2.30	1.73
$5 \text{ mg of I as KIO}_3$	9.30	3.37	1.60	1.20
CSRM				
1 mg of I as KI	10.39	4.02	1.85	1.38
3 mg of I as KI	10.07	3.40	1.72	1.12
5 mg of I as KI	10.10	3.47	1.83	1.11
1 mg of I as KIO ₃	9.50	3.44	1.55	1.16
$3 \text{ mg of I as KIO}_3$	9.65	3.41	1.66	1.17
$5 \text{ mg of I as KIO}_3$	9.16	2.84	1.55	0.91
SEM	0.209	0.166	0.060	0.063
P-value	0.746	0.444	0.332	0.409
Kind of diet				
CSM	9.58	3.87	1.81	1.29
CSRM	9.81	3.43	1.69	1.14
I source				
KI	9.71	3.69	1.75	1.19
KIO ₃	9.68	3.61	1.75	1.24
Level of I supplement, mg				
1	9.91	3.89	1.74	1.27
3	9.64	3.64	1.79	1.22
5	9.54	3.42	1.72	1.15
Source of variation, <i>P</i> -value				
Kind of diet	0.587	0.185	0.349	0.244
I source	0.946	0.807	0.957	0.669
I level	0.760	0.525	0.898	0.736
Kind of diet $\times$ I source	0.099	0.344	0.070	0.167
Kind of diet $\times$ I level	0.932	0.961	0.867	0.670
I source $\times$ I level	0.279	0.086	0.071	0.089
Kind of diet $\times$ I source $\times$ I level	0.569	0.388	0.300	0.440

¹All differences among means were nonsignificant.

 2 CSM = corn-soybean mixture; CSRM = corn-soybean-rapeseed meal mixture; TG = triglycerides; CHOL = cholesterol; HDL = high-density lipoprotein; LDL = low-density lipoprotein.

sons of egg sampling, but statistically it was confirmed only after 3 mo of the experiment (Table 9). Iodine concentration in eggs of hens receiving rapeseed meal in the ration was 522 and 673  $\mu$ g/kg, whereas in eggs of hens that did not receive rapeseed meal it was 671 and 841  $\mu$ g/kg at the first and second time of sampling, respectively. The KI was more effective source of I than  $KIO_3$  in accumulation of this element in wet egg content, especially after 5 mo of treatment (816 vs. 698  $\mu$ g/ kg). Increasing I concentration in the diet of hens led to an increase in I concentrations in egg content and this was confirmed statistically (P < 0.01). Data obtained in mo 3 of the experiment are difficult to interpret due to significant interactions between some experimental factors, but after 5 mo of the experiment, all interactions were nonsignificant. Iodine content in animal origin products, including eggs, depends on many factors [e.g., the kind and level of I supplements (Marcilese et al., 1968; Perry et al., 1989; Yue and Kang, 1995; Lewis, 2004; Röttger et al., 2012; Saki et al., 2012), the kind of diet (Wemheuer and Paufler, 1993; Tripathi et al., 2001a; Lichovnikova and Zeman, 2004), the period of additive application (Nair et al., 1998), genetic characteristics of animals (Ryś et al., 1997), animal age, hen laying rates, as well as on the chemical methods used

for the determination of I content in biological materials (Knapp et al., 1998; Gasior and Szczypuła, 2010)]. These dependences have been confirmed by numerous authors, but there have been a range of often differing results concerning rising I concentrations in whole egg, yolk, or albumen after enrichment of diets with I (Ryś et al., 1997; Kaufmann et al., 1998; Krupová et al., 1998; Dobrzański et al., 2001; Opaliński et al., 2012). Similar to our data, Yalçin et al. (2004) reported that increasing I levels in hen diets from 3 to 24 mg/kg caused a clear increase in I content both in egg albumen and egg yolk. Röttger et al. (2012) also assayed progressive increases in I accumulation in yolk  $(425-3,373 \ \mu g/s)$ kg) and in albumen (14–124  $\mu$ g/kg), as well as in whole egg (144–1,304  $\mu$ g/kg) as a response of laying hens to increasing I supplementation of diets.

In conclusion, the introduction of an I supplement from 1 to 5 mg/kg into the diet of hens enables the significant enrichment of the egg content without breaching European Union regulations. At a mean wet egg content of about 50 g and a supply of 5 mg of I/kg of feed, a consumer may consume in one egg about 51 to 63  $\mu$ g of I compared with the average recommended daily I consumption per adult person of about 160  $\mu$ g (WHO, 2001). Physiological indices determined after

Table 9. Iodine accumulation (µg/kg) in wet egg content after 3 and 5 mo of the experiment  1 

	I concer	ntration
Item	After 3 mo	After 5 mo
Treatment		
CSM		
1 mg of I as KI	$327^{ABC}$	$466^{AB}$
3 mg of I as KI	$490^{AB}$	$995^{\mathrm{CD}}$
5 mg of I as KI	$1,168^{D}$	$1,325^{CE}$
1  mg of I as KIO ₃	$408^{ABC}$	$271^{AB}$
$3 \text{ mg of I as KIO}_3$	$630^{AE}$	$618^{A}$
$5 \text{ mg of I as KIO}_3$	$1,006^{D}$	$1,373^{E}$
CSRM		
1 mg of I as KI	$113^{\text{C}}$	$269^{AB}$
3 mg of I as KI	$634^{AE}$	$644^{\mathrm{AD}}$
5 mg of I as KI	$999^{D}$	$1,198^{CE}$
1 mg of I as KIO ₃	$192^{BC}$	$234^{\mathrm{B}}$
$3 \text{ mg of I as KIO}_3$	$322^{ABC}$	$565^{AB}$
$5 \text{ mg of I as KIO}_3$	$871^{\text{DE}}$	$1.129^{CE}$
SEM	50.97	61.01
P-value	0.000	0.000
Kind of diet		
CSM	$671^{A}$	$814^{A}$
CSRM	$522^{B}$	$673^{B}$
I source		
KI	622	$816^{A}$
KIO3	571	$698^{\mathrm{B}}$
Level of I supplement, mg		
1	$260^{\text{A}}$	$310^{A}$
- 3	$519^{B}$	$705^{B}$
5	$1.011^{\circ}$	$1.256^{\circ}$
Source of variation, <i>P</i> -value	-,	-,
Kind of diet	0.000	0.000
I source	0.167	0.003
I level	0.000	0.000
Kind of diet $\times$ I source	0.057	0.136
Kind of diet $\times$ I level	0.322	0.618
I source × I level	0.040	0.072
Kind of diet $\times$ I source $\times$ I level	0.014	0.083
A-Fac use a set	1.1 1.00	1.00

^{A–E}Means within a column marked with different superscripts differ significantly at P < 0.01.

 $^1\mathrm{CSM}=\mathrm{corn}\text{-soybean}$  mixture;  $\mathrm{CSRM}=\mathrm{corn}\text{-soybean}\text{-rapeseed}$  meal mixture.

long-term I fortification in blood serum show the correct health status of birds, which may be considered model animals for humans.

## ACKNOWLEDGMENTS

This research was performed within the project BIO-FOOD—innovative, functional products of animal origin no. POIG.01.01.02-014-090/09, cofinanced by the European Union from the European Regional Development Fund within the Innovative Economy Operational Programme 2007–2013.

#### REFERENCES

- AOAC International. 2000. Official Methods of Analysis. AOAC Int., Arlington, VA.
- Arrington, L. R., R. A. Santa Cruz, R. H. Harms, and H. R. Wilson. 1967. Effects of excess dietary iodine upon pullets and laying hens. J. Nutr. 92:325–330.
- Baker, D. H. 2004. Iodine toxicity and its amelioration. Exp. Biol. Med. (Maywood) 229:473–478.

- Brzóska, F., Z. Szybiński, and P. Strzetelski. 2005. Iodine content of feeds and the possibility increasing it in milk . Med. Weter. 61:145–149. (in Polish).
- Bürgi, H., T. H. Schaffner, and J. P. Seiler. 2001. The toxicology of iodate: A review of the literature. Thyroid 11:449–456.
- Delange, F. 1994. The disorders induced by iodine deficiency. Thyroid 4:107–128.
- Delange, F. 1998. Risks and benefits of iodine supplementation. Lancet 351:923–924.
- Delange, F. 2002. Iodine deficiency in Europe and its consequences: An update. Eur. J. Nucl. Med. Mol. Imaging 29:S404–S416.
- Dobrzański, Z., H. Górecka, G. Strzelbicka, J. Szczypel, and T. Trziszka. 2001. Study on enrichment of hen eggs with selenium and iodine. Electronic J. Polish Agricult. Univ. Animal Husbandry 4:2.
- EFSA. 2005. Opinion of the Scientific Panel on Additives and Products of Substances used in animal feed on the request from the commission on the use of iodine in feeding stuffs–Question No. EFSA–Q-2003–58. EFSA J. 168:1–42.
- EU Commission. 2005. Commission Regulation (EC) No 1459/2005 amending the conditions for authorization of a number of feed additives belonging to the group of trace elements. Off. J. Eur. Union, LL 233–8-L233/10.
- Gasior, R., and M. Szczypuła. 2010. Validation of method for determination of iodine in food and biological material. Rocz. Nauk. Zoot. 37:63–73. (in Polish).
- GfE. 1999. Empfehlungen zur Energie- und Nhrstoffversorgung der Legehennen und Masthhner (Broiler). DLG Verlag, Frankfurt, Germany.
- Gheisari, A. A., P. Ghayor, S. Eghbal-Saeid, M. Toghyani, and A. A. Najafi. 2011. Effect of different dietary levels of rapeseed meal on reproductive performance of Iranian indigenous breeder hens. Asian J. Anim. Vet. Adv. 6:62–70.
- Goldhaber, S. B. 2003. Trace element risk assessment: Essentiality vs. toxicity. Regul. Toxicol. Pharmacol. 38:232–242.
- Groppel, B., W. A. Rambeck, and J. Gropp. 1991. Iodanreicherung in Organen und Geweben von Mastküken nach Iodsupplementation des Futters. Mengen- und Spurenelemete 11. Arbeit-Stagung. Jena. 300–308.
- Haldimann, M., A. Alt, A. Blanc, and K. Blondeau. 2005. Iodine content of food groups. J. Food Compos. Anal. 18:461–471.
- He, M. L., C. Borger, M. Schuster, W. Hollwich, V. Krupova, J. Kursa, E. Bonabeze, D. Ranzland, and W. W. Rambeck. 2000. Carryover of iodine from algae to the products of pigs and laying hens. Pages 251–253 in Proc. 6th Symp. Pig and Poultry Nutrition, Lutherstadt-Wittenberg.
- He, M. L., W. Hollwich, and W. A. Rambeck. 2002. Supplementation of algae to the diet of pigs: A new possibility to improve the iodine content in the meat. J. Anim. Physiol. Anim. Nutr. (Berl.) 86:97–104.
- Hetzel, B. S., and G. F. Maberly. 1986. Iodine. Pages 139–208 in Trace Elements in Human and Animal Nutrition. Vol. 2. 5th ed. W. Mertz, ed. Academic Press, New York, NY.
- Kaufmann, S., G. Wolfram, F. Delange, and W. A. Rambeck. 1998. Iodine supplementation of laying hen feed: A supplementary measure to eliminate iodine deficiency in humans? Z. Ernahrungswiss. 37:288–293.
- Knapp, G., B. Maichin, P. Fecker, S. Hasse, and P. Schramel. 1998. Iodine determination in biological materials: Options for sample preparation and final determination. J. Anal. Chem. 362:508– 513.
- Krasnodebska-Depta, A. 2005. Chapter: Diagnostic. Pages 679–686 in Poultry Disease. M. Mazurkiewicz, ed. AWA Wrocław.
- Krupová, V., P. Kratochvil, S. Kaufmann, J. Kursa, and J. Trávníček. 1998. Metabolic effects of iodine addition to laying hens. Vet. Med. (Praha) 43:207–212.
- Lewis, P. D. 2004. Responses of domestic flow to excess iodine: A review. Br. J. Nutr. 91:29–39.
- Lichovnikova, M., and L. Zeman. 2004. The effects of a higher amount of iodine supplement on the efficiency of laying hens fed extruded rapeseed and on eggshell quality. Czech J. Anim. Sci. 49:199–203.
- Liu, N. Q., Q. Xu, X. L. Hou, P. S. Liu, Z. F. Chai, L. Zhu, Z. Y. Zhao, Z. H. Wang, and Y. F. Li. 2001. The distribution patterns

of trace element in the brain and erythrocytes in a rat experimental model of iodine deficiency. Brain Res. Bull. 55:309–312.

- Marcilese, N. A., R. H. Harms, R. M. Valsecchi, and L. R. Arrington. 1968. Iodine uptake by ova of hens given excess iodine and effect upon ova development. J. Nutr. 94:117–120.
- Mathe, D., and F. Chevallier. 1976. Effects of the thyroid state on cholesterol metabolism in the rat. Biochim. Biophys. Acta 441:155–164.
- McDowell, L. R. 1992. Iodine. Pages 225–245 in Minerals in Animal and Human Nutrition. Academic Press Inc., New York, NY.
- Moore, S., and N. H. Stein. 1963. Discussion of classical methods of acid hydrolysis. Methods Enzymol. 6:819–823.
- Nair, K. M., B. Sesikeran, S. Ranganathan, and B. Sivakumar. 1998. Bioeffect and safety of long-term feeding of common salt fortified with iron and iodine (double fortified salt) in rat. Nutr. Res. 30:163–166.
- NRC. 1994. Nutrient Requirements of Poultry. 9th rev. ed. National Academies Press, Washington, DC.
- Opaliński, S., B. Dolińska, M. Korczyński, K. Chojnacka, Z. Dobrzański, and F. Ryszka. 2012. Effect of iodine-enriched yeast supplementation of diet on performance of laying hens, egg traits, and egg iodine content. Poult. Sci. 91:1627–1632.
- Perry, G. C., P. D. Lewis, and M. J. Hannagan. 1989. Iodine supplementation from two sources and its effect on egg output. Br. Poult. Sci. 30:973–974.
- Perry, G. C., P. D. Lewis, and M. J. Hannagan. 1990. Responses of the laying hen to dietary iodine supplementation. Proc. 8th Eur. Poult. Conf. WPSA. Barcelona, 1, 384–387.
- Richter, G. 1995. Einfluss der Jodversorgung der Legehennen auf den Jodgehalt im Ei. Pages 457–464 in Proc. 15. Arbeitstagung Mengen und Spurenelemente, Friedrich-Schiler-Universitat, Jena, Germany.
- Röttger, A. S., I. Halle, H. Wagner, G. Breves, S. Dänicke, and G. Flachowsky. 2012. The effects of iodine level and source on iodine carry-over in eggs and body tissues of laying hens. Arch. Anim. Nutr. 66:385–401.
- Ryś, R., E. Weir-Konas, H. Pyska, M. Kuchta, and M. Pietras. 1997. Changes in egg iodine concentration in three hen strains in relation to iodine level in diets. Rocz. Nauk. Zoot. 24:229–242. (in Polish).
- Saki, A. A., M. A. Farisar, H. Aliarabi, P. Zamani, and M. Abbasinezhad. 2012. Iodine-enriched egg production in response to dietary iodine in laying hens. J. Agric. Technology 8:1255–1267.
- Sourgens, H., A. Frömblich-Borges, and P. H. Kemper. 1986. Iodine and autoregulation of thyroid gland function: Inhibition of secretion of thyroid hormones in the rat. 5. Spurenelement-Symposium, Jod. J. Boden, Pflanze, Tier, Mensch, Friedrich-Schiller-Universitat, Jena. 53–59.
- StatSoft Inc. 2012. Statistica Data Analysis Software system, ver. 9. StatSoft Inc., Tulsa, OK.

- Szybinski, Z., F. Delange, A. Lewinski, J. Podoba, M. Rybakowa, R. Wasik, L. Szewczyk, B. Huszno, F. Golkowski, E. Przybylik-Mazurek, M. Karbownik, T. Zak, J. Pantoflinski, M. Trofimiuk, and I. Kinalska. 2001. A programme of iodine supplementation using only iodised household salt is efficient—The case of Poland. Eur. J. Endocrinol. 144:331–337.
- Trávníček, J., P. Kratochvil, and V. Kroupova. 1997. Die Entwicklung der Embryonen iodreicher Bruteier. 17. Arbeitstagung "Mengen und Spurenelemente", Friedrich-Schiller-Universitat, Jena. 821–827.
- Trávníček, J., V. Kroupová, and J. Kursa. 2000. The effects of excessive iodine intake on the activity of leukocytes and the level of plasmatic proteins in laying hens. Sci. Agric. Biohemica. 4:273–284.
- Tripathi, M. K., I. S. Agrawal, S. D. Sharma, and D. P. Mishra. 2001a. Effect of substitution of soybean meal with treated or untreated high glucosinolate mustard (*Brassica juncea*) meal on intake, digestibility, growth performance and body composition of calves. Anim. Feed Sci. Technol. 94:137–146.
- Tripathi, M. K., I. S. Agrawal, S. D. Sharma, and D. P. Mishra. 2001b. Effect of untreated, HCl treated or copper and iodine supplemented high glucosinolate mustard (*Brassica juncea*) meal on nutrient utilization, liver enzymes, thyroid hormones and growth of calves. Anim. Feed Sci. Technol. 92:73–85.
- Van Middlesworth, L. 1996. Iodinated thyroid proteins which accumulate during iodine deficiency. 16. Arbeitstagung" Mengen und Spurenelemente". Friedrich-Schiller-Universitat, Jena. 920–921.
- Wemheuer, K. W., and S. Paufler. 1993. Wirkung von Kraftfuttergaben mit hohem Anteil an Rapsextrakttionschrot (30%, 00-Raps) auf die Schilddrusenfunktion und Milchjodidgehalt bei Küken. 13. Arbeitstagung "Mengen und Spurenelemente", Friedrich-Schiller-Universitat, Jena. 223–230.
- Wheeler, R. S., and E. Hoffman. 1950. The etiology of goiter in chicks from thyroprotein-fed hens; negative role of inorganic iodine. Poult. Sci. 29:306–307.
- WHO, UNICEF, ICCIDD. 2001. Assessment of iodine deficiency disorder and monitoring their elimination. Page 107. WHO, Geneva, Switzerland.
- WPSA. 1989. European Table of Energy Values for Poultry Feedstuffs. 3rd ed. 11–28. Beekbergen, the Netherlands.
- Yalçin, S., Z. Kahraman, S. S. Yalçin, and H. E. Dedeoglu. 2004. Effects of supplementary iodine on the performance and egg traits of laying hens. Br. Poult. Sci. 45:499–503.
- Yue, W. B., and J. Q. Kang. 1995. Effects of a high-iodine diet on performance, metabolism and egg quality of layers. Acta Vet. Zootech. Sin. 26:18–22.
- Zhang, W., A. Mnastsakanov, R. Hower, H. Cantor, and Y. Wang. 2005. Urinary iodine assays and ionophore based potentiometric iodide sensors. Front. Biosci. 10:88–93.